

(19)



Europäisches Patentamt

European Patent Office

Office européen des brevets



(11)

**EP 0 341 413 B2**

(12)

**NEW EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention  
of the opposition decision:  
**17.05.2000 Bulletin 2000/20**

(51) Int. Cl.<sup>7</sup>: **A61M 1/36**, B01D 15/00

(45) Mention of the grant of the patent:  
**27.10.1993 Bulletin 1993/43**

(21) Application number: **89105890.1**

(22) Date of filing: **04.04.1989**

(54) **An adsorber module and adsorber apparatus for whole blood treatment**

Adsorbermodul sowie Adsorberapparat zur Behandlung von Vollblut

Module et appareil d'adsorption pour le traitement du sang total

(84) Designated Contracting States:  
**DE FR GB IT**

(30) Priority: **04.04.1988 JP 8127688**

(43) Date of publication of application:  
**15.11.1989 Bulletin 1989/46**

(73) Proprietor:  
**ASAHI MEDICAL Co., Ltd.**  
**Chiyoda-Ku Tokyo (JP)**

(72) Inventors:  
• **Kuroda, Toru**  
**Oita-shi Oita-ken (JP)**  
• **Tohma, Norio**  
**Ohno-gun Oita-ken (JP)**

(74) Representative:  
**Strehl Schübel-Hopf & Partner**  
**Maximilianstrasse 54**  
**80538 München (DE)**

(56) References cited:

<b>EP-A- 0 034 304</b>	<b>EP-A- 0 056 977</b>
<b>EP-A- 0 082 345</b>	<b>EP-A- 0 103 816</b>
<b>EP-A- 0 143 369</b>	<b>EP-A- 0 155 534</b>
<b>EP-A- 0 230 247</b>	<b>WO-A-80/02805</b>
<b>WO-A-85/03011</b>	<b>DE-C- 2 833 493</b>
<b>FR-A- 2 510 412</b>	<b>JP-A- 60 080 461</b>
<b>JP-A- 60 155 109</b>	<b>JP-A- 61 247 464</b>
<b>US-A- 3 723 306</b>	

- "Membranen und Membranprozesse", Staude, VCH Verlagsgesellschaft mbH, 1992, p. 30
- "Mikrofiltration mit Membranen", Ripperger, VCH Verlagsgesellschaft mbH, 1992, pp. 70,71

**EP 0 341 413 B2**

## Description

### Background of The Invention

#### 5 Field of The Invention

**[0001]** The present invention relates to an adsorber module for removing malignant components of whole blood by absorption and an adsorber apparatus containing the adsorber module. More particularly, the present invention is concerned with an adsorber module for removing malignant components of whole blood by absorption comprising a casing provided with a blood introduction means and a blood withdrawal means and a plurality of porous hollow fibers accommodated in the casing, wherein each porous hollow fiber comprises a membranous porous resin matrix having pores there within and openings on both surfaces thereof, the pores cooperating with the openings to form throughpaths running between both the surfaces of the resin matrix and a plurality of ligands linked to the overall surface of the porous resin matrix. Also, the present invention is concerned with an adsorber apparatus comprising the above-mentioned adsorber module, a blood introducing passage means having one end fluid-tightly connected to the blood introduction of the module and a blood withdrawing passage means having one end fluid-tightly connected to the blood withdrawal means of the module.

**[0002]** By the use of the adsorber module and apparatus according to the present invention, it is feasible to accomplish effective, efficient treatment of whole blood without the danger of blood coagulation and hollow clogging so that the malignant components of blood can be effectively removed by adsorption.

### Discussion of Related Art

**[0003]** In recent years, as a result of the remarkable progress of medicine (especially internal medicine), hematology, immunology, clinical inspection, etc., various malignant components have been isolated from patient blood and plasma and identified, which are believed to have close relation with the cause and advance of diseases. For example, such malignant components include autoantibodies and immune complexes against autoimmune diseases, such as rheumatoid arthritis, systemic lupus erythematosus, hyperlipemia and the like. Also, such malignant components include low or very low density lipoprotein against familial hyperlipidemia and substances of middle or low molecular weights which are increased by liver diseases.

**[0004]** Accordingly, efforts have been made in the art for developing an advantageous method by which the above-mentioned malignant component can be effectively removed from whole blood to thereby relieve or entirely cure the diseases caused by the component.

**[0005]** The proposals which have hitherto been made in line with the above-mentioned efforts are classified into (1) the one in which use is made of an active carbon or an active carbon having the surface thereof covered with a hydrophilic polymer material to remove the malignant component of plasma from whole blood, (2) the one in which whole blood is separated into a blood cell portion and a plasma portion, followed by removal of the malignant component from the plasma portion using an adsorbent (see, for example, Japanese Patent Application Publication Specification No. 55-22107/1980), and (3) the one in which the plasma separated from whole blood according to a method as mentioned in item (2) above is passed through a filter so as to remove the malignant component of high molecular weight from the plasma (see, for example, Japanese Patent Application Publication Specification No. 63-62/1988).

**[0006]** However, the proposals of items (2) and (3) above have disadvantages in that since separation of plasma from whole blood must be performed prior to adsorption or filtration, complicated apparatus is necessary and the operation is not easily accomplished. Moreover, in these proposals, since the priming volume (total of the inner volumes of apparatus components inclusive of a plasma separator, an adsorber or filter and conduits which are filled with blood) has inevitably been large, the volume of the body fluid taken from a patient has also been large to the detriment of the patient.

**[0007]** The proposals of item (1) above permit simpler apparatus and easier operation than those employed in the proposals in items (2) and (3) above, since whole blood treatment is performed without plasma separation. However, active carbon has poor selectivity for adsorption. Further, the pore size of active carbon is so small that the malignant component of the plasma cannot be adsorbed to a desirable extent.

**[0008]** A process and a device for the removal of substances from whole blood are further disclosed in WO-80/02805. This device comprises a semipermeable microporous membrane in the form of a flat membrane or of hollow fibers, which have a micro-biologically active material asymmetrically immobilized in and on the surface of the membrane which faces away from the whole blood. The biologically active materials considered there are antibodies, antigens, enzymes, protein A, killed bacteria or fragments thereof, which show strong antigenicity.

**[0009]** Due to the fact that the active material is present only in the outer surface of the membranes, such as the hollow fibers, the adsorbing capacity of the hollow fibers is rather low. Since further the biologically active substances

having antigenic activity are normally rather unstable, sterilization is likely to cause the ligand to have a disadvantageously lowered activity or to cause the ligand to be detached from the fiber, thereby bringing about serious problems with respect to safety.

**[0010]** It is, therefore, an object of the present invention to provide an adsorber module having increased adsorptive capacity for malignant substances by which it is easy to accomplish effective, efficient removal of the malignant components of whole blood without the danger of blood coagulation and hollow clogging so that the malignant components of the blood can be effectively removed by adsorption.

**[0011]** It is another object of the present invention to provide an adsorber module which is not adversely affected by sterilization and can be used in whole blood treatment with safety.

**[0012]** It is a further object of the present invention to provide an adsorber apparatus containing an adsorber of the above character in which the priming volume is small.

**[0013]** It is still a further object of the present invention to provide an adsorber apparatus which is simple in construction and easy in operation.

#### Summary of The Invention

**[0014]** With a view toward developing an effective device for removing the malignant component of whole blood which can achieve the above objects, the present inventors have made extensive and intensive studies. As a result, it has unexpectedly been found that the desired objects can be achieved by an adsorber module for removing malignant components of whole blood by adsorption, comprising:

a casing provided with a blood introduction means (2) and a blood withdrawal means (3), and a plurality of porous hollow fibers (1) substantially equal in length arranged substantially in parallel relationship and bonded together at both end portions thereof to form a bundle, each porous hollow fiber of said bundle having openings at both terminal ends thereof, said bundle being disposed in said casing along the length of said casing, said both end portions of the hollow fibers of said bundle being fluid-tightly connected to said blood introduction means and said blood withdrawal means, respectively, thereby establishing communication between said blood introduction means and said blood withdrawal means through said bundle of hollow fibers, each porous hollow fiber comprising a membranous porous resin matrix having pores therewithin and openings on both surfaces thereof, said pores cooperating with said openings to form throughpaths running between both the surfaces of said resin matrix, and a plurality of ligands (8) attached to the overall surface of said porous resin matrix, each of said ligands being capable of selectively interacting with the malignant components to be adsorbed on said hollow fibers, said ligands having low antigenicity such that when the ligands are in contact with blood corpuscles, the ligands do not cause hemolysis of erythrocyte, sensitization of leukocyte, or adherence and/or aggregation of platelet, said porous resin matrix having an average pore diameter of from 0.005 to 3  $\mu\text{m}$ , said porous hollow fibers having an average effective length (L mm) and an average inner diameter (D mm) which satisfy the inequalities

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000, \text{ and } 150 \mu\text{m} \leq D \leq 400 \mu\text{m}$$

said average effective length being defined as an average of the lengths of said porous hollow fibers minus the lengths of said both end portions of said porous hollow fibers at which the fibers are bonded together and fluid-tightly connected to said whole blood introduction means and said blood withdrawal means, respectively.

**[0015]** The invention further relates to an adsorber apparatus for removing malignant components of whole blood by adsorption which comprises the above-defined adsorber module.

#### Brief Description of The Drawings

**[0016]** In the drawings:

Fig. 1 is a diagrammatic cross-sectional view of one form of an adsorber module according to the present invention; Fig. 2 is an enlarged, diagrammatic cross-sectional view of a portion of a preferred form of hollow fiber used in the present invention, illustrating a plurality of ligands linked to the porous resin matrix of the hollow fiber. Fig. 3 is a diagrammatic view of one form of an adsorber apparatus according to the present invention; Fig. 4 is a diagrammatic view of another form of an adsorber apparatus according to the present invention; and

Fig. 5 is a diagrammatic view of a comparative adsorber apparatus.

[0017] In Figs. 1 through 5, like parts or portions are designated by like numerals.

## Detailed Description of The Invention

[0018] Referring now to Fig. 1, there is shown a schematic cross-sectional view of one form of adsorber module of the present invention which has opening 4 in the wall of the casing. The adsorber module comprises a casing provided with blood introduction means 2 and blood withdrawal means 3; and a plurality of porous hollow fibers 1 substantially equal in length which are arranged substantially in parallel relationship and bonded together at both end portions thereof to form a bundle. Each porous hollow fiber 1 of the bundle has openings at both terminal ends thereof. The bundle is accommodated in the casing along the length of the casing. The both end portions of hollow fibers 1 of the bundle are fluid-tightly connected to blood introduction means 2 and blood withdrawal means 3, respectively, thereby establishing communication between blood introduction means 2 and blood withdrawal means 3 through the bundle of hollow fibers 1.

[0019] The porous hollow fiber employed in the adsorber module of the present invention comprises a membranous porous resin matrix having pores which open at least at the inner wall of the hollow fiber and a plurality of ligands linked or attached to the overall surface of the porous resin matrix. The terminology "overall surface" as used herein means all of the inner and outer surfaces of the hollow fiber membrane and the surfaces defined by the walls of open pores present within the membranous porous resin matrix.

[0020] In Fig. 2, there is shown a schematic cross-sectional view of a preferred form of porous hollow fiber used in the present invention. Porous hollow fiber 1 shown in Fig. 1 comprises membranous porous resin matrix (also designated numeral 1 in Fig. 2) and a plurality of ligands 8 linked to the overall surface of the porous resin matrix. The resin matrix 1 has through-paths running between both surfaces 5 and 6 of the resin matrix. As mentioned above, ligands 8 are linked or attached to the overall surface of matrix 1, that is, inner wall surface 5, outer wall surface 6 and pore wall surface 7.

[0021] In the present invention, the membranous porous resin matrix of each hollow fiber have pores therewithin and openings on both surfaces thereof, which pores cooperate with the openings to form through-paths running between both the surfaces of the resin matrix.

[0022] The average pore diameter of the porous resin matrix is not particularly limited in the present invention. The pore diameter is generally selected taking into account the size and shape of the malignant component to be removed by the porous hollow fiber membrane. It is preferred that the hollow fiber have a pore diameter at which the plasma containing the malignant component is freely passed through the membrane so that the inner wall surfaces of the pores are so large as to permit unlimited contact of the plasma containing a malignant component with the ligands linked to the porous resin matrix. The average pore diameter is determined as follows. The pore diameter and pore volume of the membrane are measured by means of a mercury porosimeter. The logarithm of the diameter is plotted as the abscissa and the pore volume is plotted as the ordinate to give a pore diameter distribution curve. Thus, the total pore volume is defined by the abscissa and the pore diameter distribution curve. A vertical line can be drawn in parallel to the ordinate so that the total pore volume is halved. The value of the pore diameter on the abscissa at its point crossed by the above-mentioned vertical line is referred to as the "average pore diameter". The average pore diameter is in the range of from 0.005 to 3  $\mu\text{m}$ , preferably from 0.01 to 2  $\mu\text{m}$ , more preferably from 0.02 to 1  $\mu\text{m}$ .

[0023] The area of the overall surface of the porous resin matrix is not critical in the present invention. However, the area of the overall surface is generally at least 5  $\text{m}^2/\text{g}$ , preferably at least 10  $\text{m}^2/\text{g}$ , more preferably at least 15  $\text{m}^2/\text{g}$ . The area of the overall surface is determined as follows. The nitrogen gas adsorption amount of the matrix is measured using a BET (S. Brunauer - P<sup>o</sup>H. Emmett - E. Taylor) type surface area measuring apparatus, and the overall surface is determined from the nitrogen gas adsorption amount according to the customary one-point method.

[0024] In the present invention, the porous hollow fibers have an average effective length (L mm) and an average inner diameter (D mm) which satisfy the inequalities:

$$L/D^2 (\text{mm}^{-1}) \geq 2000 \text{ and } 150 \mu\text{m} \leq D \leq 400 \mu\text{m}$$

[0025] The average effective length is defined as an average of the lengths of the porous hollow fibers minus the lengths of both end portions of the porous hollow fibers which both end portions are bonded together and fluid-tightly connected to blood introduction means 2 and blood withdrawal means 3, respectively.

[0026] In the adsorber module of the present invention, each porous hollow fiber is adapted to allow whole blood to flow within the hollow thereof along the length of the fiber while causing the blood to be contacted with the wall surface of each hollow fiber, so that the malignant components of the blood is adsorbed on the hollow fiber by the interaction between the ligands and the malignant components. Since a large surface is effectively utilized for whole blood to be

contacted with the ligands linked or attached to surface of the porous resin matrix, effective, efficient removal of the malignant components of whole blood can be attained.

**[0027]** When the membranous porous resin matrix of each hollow fiber have pores therewithin and openings on both surfaces thereof, which pores cooperate with the openings to form through-paths running between both the surfaces of the resin matrix, the volume ratio of erythrocyte to whole blood (hematocrit) is generally higher at portions of the hollow of the porous hollow fiber which are on the side of the blood withdrawal means than at portions of the hollow of the porous hollow fiber which are on the side of the blood introduction means. This is attributed to permeation of the plasma through the wall of the hollow fiber from the inside of the fiber to the outside of the fiber. The above-mentioned increase in the volume ratio of erythrocyte brings about an increase in the viscosity of the blood flowing through the hollow of the porous hollow fiber. The increase in blood viscosity produces an advantageous pressure by which the above-mentioned permeation of blood plasma is promoted. On the other hand, in the adsorber module on the side of the blood withdrawal means, plasma which has been passed through the hollow fiber membrane and has entered the interstices extending between the outer wall of the porous hollow fibers and the inner wall of the casing and between the outer walls of the individual hollow fibers is returned to the hollow of the porous hollow fiber by the action of the increased pressure of the plasma in the outside of the hollow fiber. The plasma joins the blood having an increased content of erythrocyte in the hollow of the porous hollow fiber, and then the resultant blood is discharged from the adsorber module through the blood withdrawal means. When the plasma first passes from the inside to the outside of the hollow fiber through the hollow fiber wall membrane, a great part of the malignant component is adsorbed. And, when the plasma then passes from the outside to the inside of the hollow fiber through the hollow fiber wall membrane, the malignant component is almost completely removed.

**[0028]** The plasma having most the malignant component removed by first passing it from the inside to the outside of the hollow fiber through the wall membrane, may be discharged from the casing through at least one opening provided for withdrawing the plasma. The plasma discharged from the casing flows through a conduit connected to the opening, and then joins the blood flowing through a conduit which is connected to the blood withdrawal means of the casing, as described later.

**[0029]** From the viewpoint of minimizing the priming volume as well as increasing the malignant component-separating efficiency by the passage of plasma through the hollow fiber membrane, it is preferred that the inner diameter of the porous hollow fiber be as small as possible on the condition that blood can flow through the hollow of the porous hollow fiber. Specifically, the inner diameter of the porous hollow fiber is 150  $\mu\text{m}$  to 400  $\mu\text{m}$ .

**[0030]** In the present invention, it is preferred that the wall thickness of the porous hollow fiber be as large as possible as long as the efficiency for passing plasma through the hollow fiber membrane is not practically impaired. By increasing the wall thickness, the total surface area of the pore wall of the porous hollow fiber is increased. Specifically, the wall thickness of the porous hollow fiber is preferably 10  $\mu\text{m}$  or more, more preferably 50  $\mu\text{m}$  to 500  $\mu\text{m}$ , and most preferably 50  $\mu\text{m}$  to 200  $\mu\text{m}$ .

**[0031]** The membranous porous resin matrix to be used for preparing the present porous hollow fiber may be prepared from a hydrophilic material, such as cellulose, a cellulose derivative, a water-insoluble polyvinyl alcohol and a copolymer of ethylene and vinyl alcohol or a hydrophobic material, such as a polyolefin (e.g., polyethylene or polypropylene), a polysulfone and a polytetrafluoroethylene. It should be noted that when a hydrophobic material is used, since an aqueous solution is not readily passed through a porous hollow fiber made of the hydrophobic material, it is generally necessary to render the surface of the porous hollow fiber hydrophilic, e.g., by coating the surface of the porous hollow fiber with a hydrophilic material or by subjecting the surface of the porous hollow fiber membrane to chemical treatment or plasma treatment.

**[0032]** From the viewpoint of facilitating the linking of the ligands which interact with the malignant component at the surface of the membranous porous resin matrix, it is preferred that the matrix have functional groups to which the ligands are readily bonded, such as a hydroxyl group, an amino group, a carboxyl group and a thiol group. However, since ligands can be linked to the surface of the porous hollow fiber by physical adsorption, embedding, insolubilizing precipitation onto the matrix surface and the like, the porous resin matrix does not necessarily have to possess may such functional groups.

**[0033]** There is no particular limitation with respect to the method for forming a porous hollow fiber. That is, a porous hollow fiber can be formed by a customary method, such as wet spinning, dry spinning, melt spinning or the like. It is preferred to prepare a porous hollow fiber by a stretching perforation method in which a crystalline polymer is spun, e.g., by melt spinning, and subjected to cold stretching to cause cleavage among crystalline lamellae of the polymer and then subjected to hot stretching to attain an expansion of the cleavage. This is because controlling of pore diameters of the porous resin matrix of hollow fiber is easy and because the mechanical strengths of the obtained hollow fiber are high irrespective of the presence of a large number of pores. It is also preferred to prepare a porous hollow fiber from a polyolefin by the stretching perforation method from the viewpoint of the chemical resistance of the matrix of the hollow fiber during the reaction for linking ligands to the surface of the membranous porous resin matrix of hollow fiber.

**[0034]** The ligands may be linked to the surface of the membranous porous resin matrix by any of the customary

methods. For example, the ligands are bonded to the surface of the membranous porous resin matrix by, e.g., covalent bonding or ionic bonding. Also, the ligands may be bonded to the surface of the porous matrix by physical adsorption, embedding, insolubilizing precipitation onto the matrix surface or the like. Of these methods, from the viewpoint of avoiding the elution of ligands, it is preferred to use a method in which the ligands are bonded to the surface of the matrix by covalent bonding. Specifically, the conventional carrier activating method and the conventional ligand linking method, which are utilized in the preparation of an immobilized enzyme or in an affinity chromatography, may be employed. Representative examples of carrier activating methods include methods using a cyanogen halogenide, periodic acid, a crosslinking agent or an epoxide. In the carrier activating method, substitution and/or addition of active hydrogen-containing nucleophilic groups, such as an amino group, a hydroxyl group, a carboxyl group and a thiol group, to the reactive groups on the surface of the porous resin matrix is performed by chemical modification with, e.g., a cyanogen halogenide, such as CNBr. However, the carrier activating methods are not restricted to the above-mentioned methods.

**[0035]** "Ligand" used herein means a substance which selectively interacts with a malignant component to be adsorbed on the hollow fiber, which is contained in blood plasma.

**[0036]** Representative examples of ligands include an amino acid, a peptide, a protein, a glycoprotein, a monosaccharide, an oligosaccharide, a polysaccharide, a lipid, an antigenic substance, a nucleic acid, a nonprotein organic compound and an inorganic compound. Representative examples of proteins and glycoproteins include an antibody, a complement, a protein relating to blood coagulation and an enzyme.

**[0037]** However, it is necessary that the ligands exhibit no or only weak toxicity even if it is eluted into blood, and that when the ligands are in contact with blood corpuscles, the ligands do not cause hemolysis of erythrocyte, sensitization of leukocyte or adherence and/or aggregation of platelet. In this connection, when a peptide, a protein and a glycoprotein have a low weight average molecular weight, they have a low antigenicity. Therefore, with respect to a peptide, a protein and a glycoprotein, it is preferred that they have a low weight average molecular weight, although there is generally no particular restriction with respect to the molecular weight of the ligands. From these viewpoints, preferred are a peptide, a protein and a glycoprotein, each of which has a preferred weight average molecular weight of not greater than  $10^4$ ; and a monosaccharide, an oligosaccharide, a polysaccharide, a lipid, a nucleic acid, a nonprotein organic compound (e.g., a polyanion), an inorganic compound, an amino acid and a peptide each having a weight average molecular weight of not greater than  $10^3$ . The weight average molecular weight of the peptide, the protein and the glycoprotein are determined by measuring a sedimentation coefficient of the solute. That is, the sedimentation coefficient of the peptide, the protein or the glycoprotein is measured according to the sedimentation velocity method using an ultracentrifuge, and the molecular weight is calculated by substituting the sedimentation coefficient measured for the  $s$  in the following Svedberg formula:

$$\text{Weight average molecular weight (Mw)} = \frac{R \cdot T \cdot s}{D(1 - \bar{v}\rho)}$$

wherein  $R$  represents the gas constant,  $T$  represents the absolute temperature,  $s$  represents the sedimentation coefficient of a solute,  $D$  represents the diffusion coefficient of a solute,  $\bar{v}$  represents the partial specific volume of a solute and  $\rho$  represents the density of a solvent.

**[0038]** Representative examples of ligands are as follows.

**[0039]** For the treatment of systemic lupus erythematosus, preferable use is made of a mononucleoside containing adenine, guanine, cytosine, uracil or thymine, an oligonucleoside or polynucleoside comprising such a mononucleoside; and a natural nucleic acid, such as DNA and RNA. These are used to remove an anti-nuclear antibody and an anti-DNA antibody which are present in blood plasma. Further, a basic compound, such as actinomycin D, may be used to remove DNA, RNA and ENA which are present in blood plasma.

**[0040]** For the treatment of the so-called immune complex diseases, such as chronic articular rheumatism, malignant tumor and systemic lupus erythematosus, a hydrophobic compound may be used to remove an immune complex.

**[0041]** For the treatment of diseases so-called autoimmune diseases, such as myasthenia, multiple sclerosis and chronic articular rheumatism, a hydrophobic compound may be used to remove an autoantibody.

**[0042]** For the treatment of hyperlipemia, use is preferably made of a polyanion, e.g., an acid polysaccharide, such as heparin and dextran sulfate; a synthetic polyanion, such as polyvinyl sulfate and polyacrylic acid. These are used to remove a low density lipoprotein and a very low density lipoprotein.

**[0043]** The ligands to be used in the present invention are not restricted to the above-mentioned ones. The ligands may be used individually or in combination.

**[0044]** When the ligands are used in combination, the bundle to be incorporated in the present adsorber module may be comprised of hollow fibers of the same kind, each having different types of ligands or of different hollow fibers which are different in the type of ligand linked thereto.

**[0045]** With respect to a hydrophobic compound and a polyanion used as ligands, a more detailed explanation will be made as follows.

**[0046]** The hydrophobic compound to be used in the present invention is a compound having a solubility in physiological saline of not greater than 100 mmole/dl (at 25 °C), preferably not greater than 30 mmole/dl (at 25 °C). When a compound having a solubility exceeding 100 mmole/dl in physiological saline is used, the compound is so hydrophilic that the compound has poor affinity for an autoantibody and an immune complex, thereby leading to an extreme lowering in the adsorbing capacity of the adsorber module. Further, since such a compound has an affinity for an albumin which is hydrophilic, the albumin is likely to be non-selectively adsorbed on the porous hollow fiber. This is disadvantageous.

**[0047]** When a hydrophobic compound having at least one aromatic ring is used as the ligand, particularly preferable results can be obtained. A hydrophobic compound having any type of aromatic ring may be advantageously used. However, it is preferred to use a hydrophobic compound having at least one aromatic ring selected from a benzene type aromatic ring, such as a benzene ring, a naphthalene ring and a phenanthrene ring; a nitrogen-containing six-membered ring, such as a pyridine ring, a quinoline ring, an acridine ring, an isoquinoline ring and a phenanthridine ring; a nitrogen-containing five-membered ring, such as an indole ring, a carbazole ring, an isoindole ring, an indolizene ring, a porphyrin ring and 2,3,2',3'-pyrrolopyrrole ring; a six-membered ring having 2 or more nitrogen atoms, such as a pyridazine ring, a pyrimidine ring, a sym-triazine ring, a sym-tetrazine ring, a quinazolin ring, a 1,5-naphthyridine ring, a pteridine ring and a phenazine ring; a five-membered ring having 2 or more nitrogen atoms, such as a pyrazole ring, a 1,2,4-triazole ring, a 1,2,3-triazole ring, a tetrazole ring, a benzimidazole ring, an imidazole ring and a purine ring; an oxygen-containing aromatic ring, such as a norharman ring, a perimidine ring, a benzofuran ring, an isobenzofuran ring, and dibenzofuran ring; a sulfur-containing aromatic ring, such as a benzothiophene ring, a thienothiophene ring and a thiepine ring; an oxygen-containing heterocyclic aromatic ring, such as, an oxazole ring, an isooxazole ring, a 1,2,5-oxadiazole ring and a benzoxazole ring; a sulfur-containing heterocyclic aromatic ring, such as a thiazole ring, an isothiazole ring, a 1,3,4-thiadiazole ring and a benzothiazole ring; and derivatives thereof. Of these compounds, a compound having an indole ring, such as tryptamine, provides particularly preferred results. This is probably because the hydrophobic nature and rigidity of the molecule of such compounds promote the bonding of the compounds with an autoantibody and an immune complex.

**[0048]** It has been found that a hydrophobic amino acid and a derivative thereof which are practically harmless and inexpensive are extremely advantageous to remove an autoantibody and an immune complex. The hydrophobic amino acid and a derivative thereof preferably have a hydrophobic parameter of 1500 cal/mol or more, which hydrophobic parameter is defined by Tanford and Nozaki in Journal Of American Chemical Society 184, page 4240 (1962) and Journal Of Biological Chemistry 246, page 2211 (1971). Also, the hydrophobic amino acid and a derivative thereof preferably have a solubility in physiological saline of 100 mmole/dl or less (at 25 °C). When the solubility exceed 100 mmole/dl, the hydrophobic interaction between the hydrophobic amino acid and a derivative thereof and a malignant component becomes weak, thereby causing the adsorber module to have poor adsorbing ability.

**[0049]** Representative examples of hydrophobic amino acids and derivatives thereof include lysine, valine, leucine, tyrosine, phenylalanine, isoleucine, tryptophan and derivatives thereof.

**[0050]** Of these hydrophobic amino acids and derivatives thereof, phenylalanine, tryptophan and derivatives thereof are particularly preferred. There is no particular limitation with respect to the optical isomerism of the hydrophobic amino acid. That is, the hydrophobic amino acid may be dextrorotatory or levorotatory.

**[0051]** The weight average molecular weight of the hydrophobic compound to be used in the present invention is preferably 10,000 or less, more preferably 1,000 or less. The weight average molecular weight of the hydrophobic compound is determined in substantially the same manner as in the measurement of the weight average molecular weights of the peptide, the protein and the glycoprotein. When the hydrophobic compound has the above-mentioned preferable weight average molecular weight, the hydrophobic compound is readily linked to the membranous porous resin matrix of hollow fiber, and the hydrophobic compound-linked porous hollow fiber has good storage stability as compared to that of a natural polymer, such as protein S (weight average molecular weight: 42,000). Even if a hydrophobic compound having a weight average molecular weight of 10,000 or less is eluted from a porous hollow fiber into blood, since the antigenicity of the hydrophobic compound is negligibly low, there arises no safety problem for a living body. The hydrophobic compound having a weight average molecular weight of 10,000 or less is not deactivated by sterilization.

**[0052]** The term "polyanion" used herein means a polymer which has a weight average molecular weight of 600 or more and which has functional groups capable of producing a negative charge in a body fluid, such as a sulfonic group, a carboxyl group and a phosphoric group. The weight average molecular weight of the polyanion is determined in substantially the same manner as in the measurement of the weight average molecular weights of the peptide, the protein and the glycoprotein. The polyanion is preferably in the form of a straight chain polymer so as to capture a malignant component effectively. The weight average molecular weight of the polyanion is preferably in the range of 600 to  $10^7$ , more preferably 1,000 to  $5 \times 10^6$ , most preferably  $2,000 \times 10^6$ . Since a low density lipoprotein and a very low density lipoprotein, which are malignant components, have a diameter as large as  $2 \times 10^{-4}$  to  $8 \times 10^{-4}$   $\mu\text{m}$ , it is desired to use a

polyanion having the above-mentioned weight average molecular weight. When the polyanion has a weight average molecular weight of less than 600, the adsorbing capacity for a low density lipoprotein and a very low density lipoprotein is likely to be insufficient. It is preferred that the polyanion have at least one of the above-mentioned functional groups capable of producing a negative charge in a body fluid, such as a sulfonic group, a carboxylic group and a phosphoric group per 300, more preferably per 200, most preferably per 50 to 150 in weight average molecular weight of the polyanion. This is because strong electrostatic interaction can be attained between the polyanion and a malignant component. This weight average molecular weight of the polyanion means a weight average molecular weight which includes the molecular weight of the functional groups.

**[0053]** Representative examples of polyanions include vinyl type synthetic polyanions, such as a polyacrylic acid, a polymethacrylic acid, a polyvinyl sulfonic acid, a polyvinyl sulfuric acid, a polymaleic acid, polyfumaric acid and derivatives thereof; styrene type synthetic polyanions, such as a poly(styrenesulfonic acid) and a poly(styrenephosphoric acid); peptide type polyanions, such as a polyglutamic acid and a polyaspartic acid; nucleic acid type polyanions, such as a poly U and a poly A; synthetic polyanions, such as a polyphosphoric acid ester, a poly- $\alpha$ -methylstyrenesulfonic acid and copolymers of styrene and methacrylic acid; and polysaccharide type polyanions, such as heparin, dextran sulfate, chondroitin sulfate, alginic acid, pectin, hyaluronic acid and derivatives thereof. However, the polyanion to be used in the present invention is by no means restricted to the above-mentioned specific examples.

**[0054]** The ligands capable of interacting with a malignant component are linked to the overall surface of a membranous porous resin matrix, preferably with substantially uniform distribution. However, in some portions, the ligands may be non-uniformly linked to the surface of the membranous porous resin matrix. Further, in the surface of a membranous porous resin matrix, there may be present some portions to which the ligands are not linked, as long as the presence of such portions cause little adverse effect on the adsorbing capacity for a malignant component.

**[0055]** The distribution of the ligands linked to the inner wall surface and the outer wall surface of the membranous porous resin matrix can be observed by coloring the ligands according to the customary staining method. The distribution of the ligands linked to the wall surface of the pores of the porous resin matrix can be observed by immersing the porous fiber in an aqueous solution of a metal salt to form a salt of the ligand with the metal utilizing the interaction between the ligand and the metal, cutting off a thin section from the resultant porous fiber and subjecting the thin section to observation using an X-ray micro analyzer to detect the metal which is present in the form of the salt of the ligand with the metal linked to the wall of the pores of the porous fiber.

**[0056]** When the inner surface of the porous hollow fiber is subjected to treatment for prevention of hollow clogging by platelets and/or treatment for prevention of blood coagulation, more preferable results are obtained.

**[0057]** The porous hollow fibers are packed in a casing to prepare the adsorber module for blood treatment according to the present invention by the use of a customary method, e.g., a method for preparing an artificial kidney.

**[0058]** The adsorber module for whole blood treatment of the present invention can advantageously be constructed into an adsorber apparatus.

**[0059]** Accordingly, in another aspect of the present invention, there is provided an adsorber apparatus for removing malignant components of whole blood by adsorption comprising:

- (a) an adsorber module according to the invention, together with
- (b) a blood introducing passage means comprising a first conduit having one end fluid-tightly connected to the blood introduction means of module (a), a second conduit having a blood inlet at one end thereof, and a blood transport means disposed between and fluid-tightly connected to the other ends of the first and second conduits; and
- (c) a blood withdrawing passage means comprising a third conduit having one end fluid-tightly connected to the blood withdrawal means of module (a) and having a blood outlet at the other end thereof.

**[0060]** In the adsorber apparatus, the membranous porous resin matrix of each hollow fiber of the adsorber module has pores therewithin and openings on both surface thereof, which pores cooperate with the openings to form through-paths running between both the surfaces of the resin matrix. In this case, the casing of module (a) may further be provided with at least one opening for withdrawing the plasma which opening is fluid-tightly connected to the blood withdrawing passage means through a bypass comprising a fourth conduit having one end fluid-tightly connected to the opening, a fifth conduit having one end fluid-tightly connected to the blood withdrawing passage at its intermediate position, and a plasma transport means disposed between and fluid-tightly connected to the other ends of the fourth and fifth conduits.

**[0061]** Referring to Fig. 3, there is shown one form of adsorber apparatus of the present invention. The apparatus comprises (a) adsorber module 13 having no opening (as indicated in Fig. 1) in the casing for withdrawing the plasma; (b) a blood introducing passage means comprising first conduit 12 having one end fluid-tightly connected to module 13 at its blood introduction means 2 (not shown), second conduit 10 having blood inlet 9 at one end thereof, and blood transport means 11 (e.g., pump) disposed between and fluid-tightly connected to the other ends of first and second



conduits 12 and 10; and (c) a blood withdrawing passage means comprising third conduit 14 having one end fluid-tightly connected to module 13 at its blood withdrawal means 3 (not shown) and having blood outlet 15 at the other end thereof. Each porous hollow fiber 1 of module 13 is adapted to allow whole blood, which is introduced from blood inlet 9 through the blood introducing passage, to flow within the hollow thereof along the length of fiber 1 while causing the plasma of the blood to be contacted at least inner wall surface 5, including open pore walls, of each hollow fiber 1, so that the malignant component of the plasma is adsorbed on hollow fiber 1 by the interaction between ligand 8 and the malignant component.

**[0062]** In operation, whole blood is introduced to the adsorber apparatus from the patient (donor) through blood inlet 9 and flowed to adsorber module 13 at a predetermined flow rate through conduits 10 and 12 by means of blood transport means 11 (e.g., pump). The whole blood is treated during the flowing through the hollow fibers so that the malignant components of the blood are adsorbed on the ligands attached to the hollow fiber membranes. The blood treated with the adsorber module 13 is flowed through conduit 14 and returned to the donor through blood outlet 15. The whole blood from the donor is recycled through the same adsorber apparatus. The recycling is continued for a predetermined period of time. After the recycling purified, the whole blood of the patient becomes pure.

**[0063]** It is necessary that the membranous porous resin matrix of each hollow fiber 1 of adsorber module 13 have pores therewithin and openings on both surfaces and thereof, which pores cooperate with the openings to form through-paths running between both surfaces and of the resin matrix. When such hollow fibers are used in adsorber module 13, during the flowing of the whole blood through the hollows of the hollow fibers, the plasma permeates through the wall of each hollow fiber from the inside of the fiber to the outside of the fiber. The plasma which has been passed through the hollow fiber membrane and has entered the interstices extending between the outer wall of the porous hollow fibers and the inner wall of the casing and between the outer walls of the individual hollow fibers is returned to the hollow of the porous hollow fiber by the action of the increased pressure of the plasma in the outside of the hollow fiber. This is advantageous from the viewpoint of attaining a greater contact area between the whole blood and the ligand.

**[0064]** Referring to Fig. 4, there is shown another form of the adsorber apparatus of the present invention. In adsorber module 13, as described with reference to Fig. 2, the membranous porous resin matrix of each hollow fiber 1 of adsorber module 13 has pores 7 therewithin and openings on both surfaces 5 and 6 thereof, which pores 7 cooperate with the openings to form through-paths running between both surfaces 5 and 6 of the resin matrix. Therefore, as described above, when the whole blood is allowed to flow within the hollows along fibers 1, the plasma of the blood is caused to be selectively passed from the inside to the outside and, in turn, from the outside to the inside of each hollow fiber 1 reciprocally through the walls of hollow fibers 1. In the apparatus of Fig. 4, the casing of adsorber module 13 is further provided with at least one opening for withdrawing the plasma. The opening is fluid-tightly connected to blood withdrawing passage means (19 and 20) through a bypass comprising fourth conduit 16 having one end fluid-tightly connected to the opening, fifth conduit 18 having one end fluid-tightly connected to blood withdrawing passage (19 and 20) at its intermediate position, and plasma transport means 17 (e.g., pump) disposed between and fluid-tightly connected to the other ends of fourth and fifth conduits 16 and 18.

**[0065]** In operation, whole blood is introduced in the adsorber apparatus from blood inlet 9 and flowed to adsorber module 13 at a predetermined rate through conduits 10 and 12 by means of blood transport means 11 (e.g., pump). In adsorber module 13, the blood is allowed to flow through the hollows of hollow fibers 1, and the plasma of the blood is allowed to pass through the membrane of each hollow fiber. The thus separated, purified plasma is flowed to conduit 20 through a bypass, i.e., through conduits 16 and 18 by means of blood transport means 17 (e.g., pump) at a flow rate which is lower than the flow rate of the blood flowing from conduits 10 and 12 into adsorber module 13 by means of pump 11. On the other hand, the cell-enriched blood which has not been passed through the bypass is flowed to conduit 20 through conduit 19. In conduit 20, the purified plasma is combined with the cell-enriched blood and the resultant mixture is returned to the donor through blood outlet 15. The blood from the patient is recycled through the same adsorber apparatus. The recycling is continued for a predetermined period of time. After the recycling, the whole blood of the patient becomes pure.

**[0066]** As described, the adsorber module of the present invention can easily be constructed into an adsorber apparatus which can be practically employed for treatment of whole blood. With this apparatus, whole blood can be effectively, efficiently treated without the danger of blood coagulation and hollow clogging, whereby the malignant components of the whole blood can be effectively removed by adsorption.

**[0067]** As described above, the adsorber module and the adsorber apparatus of the present invention is very effective for removing malignant components of whole blood by adsorption, but they also can be effectively used for removing the malignant components of plasma separated from whole blood by means of a conventional plasma separator.

#### Detailed Description of The Preferred Embodiments

**[0068]** The present invention will now be described in more detail with reference to the following Examples, which should not be construed to be limiting the scope of the present invention.

## Example 1

**[0069]** A high-density polyethylene (HI-ZEX 2208J, a product of Mitsui Petrochemical Co., Japan) having a density of 0.968 g/cm<sup>3</sup> and a melt index, as measured in accordance with ASTM D1238, of 5.5 is extruded from an annular hollow fiber spinning nozzle having an annular orifice outside diameter of 35 mm and an annular orifice inside diameter of 27 mm (slit width: 4 mm) at a winding temperature of 150 °C and at a polymer extrusion rate of 16 g/min and a winding rate of 200 m/min. The thus obtained hollow fiber is subjected to annealing at 115 °C for 2 hours. The annealed hollow fiber is then cold-stretched at room temperature at a stretching ratio (times) (the ratio of the length of the stretched hollow fiber to the length of the hollow fiber before stretching, expressed by times) of 1.33 times by passing the annealed hollow fiber through stretching rolls arranged to provide a stretching path of 200 mm. Then, the cold-stretched hollow fiber is hot-stretched successively at 78 °C, 95 °C and 98 °C at stretching ratios at 78 °C, 95 °C and 98 °C of 3 times, 1.28 times and 1.14 times, respectively. The ratio (%) of the length of the stretched hollow fiber to the original length of the hollow fiber before cold-stretching and hot-stretching is 480 %. The thus stretched hollow fiber is heat set at 115 °C for 2 min, to thereby obtain a porous polyethylene hollow fiber.

**[0070]** The polyethylene hollow fiber is immersed in a solution of polyethylene vinyl alcohol (Soanol Z, manufactured and sold by The Nippon Synthetic Chemicals Industry Co., Ltd., Japan) in 70 % (w/w) aqueous ethanol solution, having a polyethylene vinyl alcohol concentration of 0.9 % by weight, and kept at 50 °C for 5 min in the solution. Then, the hollow fiber is taken out of the solution and dried at 55 °C for 1.5 hours. The resultant hollow fiber has an inside diameter of 340 µm, an outside diameter of 440 µm, a membrane thickness of 50 µm, an average pore diameter of 0.3 µm and a surface area of 21 m<sup>2</sup>/g.

**[0071]** The hollow fiber is cut to the length of 30 cm to obtain 2000 cut hollow fibers, and they are immersed in a mixture of 500 ml of acetone, 390 ml of epichlorohydrin and 90 ml of a 40 % (w/w) aqueous NaOH solution. The hollow fibers are subjected to ultrasonication in the mixture at 30 °C for 5 hours. Thereafter, the hollow fibers are washed with acetone and then with water, to obtain epoxy-activated polyethylene hollow fibers.

**[0072]** The epoxy-activated polyethylene hollow fibers are immersed in 1000 ml of 1 M sodium carbonate buffer (pH 9.8) containing 10.20 g of tryptophan and subjected to ultrasonication at 50 °C for 24 hours, to thereby link tryptophan to the overall surface of each of the hollow fibers. The amount of tryptophan linked as a ligand to the surface of each of the hollow fibers is 360 µmol/g (dry basis). The ligand-linked hollow fibers are dried. The dried hollow fibers are inserted in a polyethylene-made cylindrical casing so that they are arranged substantially in parallel relationship and accommodated in the casing along the length thereof. Then, both end portions of the hollow fibers and both end portions of the inner side wall of a polycarbonate-made cylindrical casing having a plasma outlet on its side wall are bonded by a centrifugal molding method using a polyurethane resin adhesive to obtain an assembly. Both the end portions of the resultant assembly are cut off to open the terminals of the hollow fibers, and an opening-having end cap is then attached to each of the end portions of the assembly as shown in Fig. 1 to provide a blood introduction means and a blood withdrawal means so that the both end portions of the hollow fibers are fluid-tightly connected to the blood introduction means and the blood withdrawal means, respectively, thereby establishing communication between the blood introduction means and the blood withdrawal means through the hollow fibers. Thus, there is obtained an adsorber module as shown in Fig. 1. The average effective length of the hollow fibers of the adsorber module is 255 mm, and the ratio of the effective length of the hollow fiber to the square of the inside diameter of the hollow fiber ( $L/D^2$ ) = 2206 mm<sup>-1</sup>.

**[0073]** Using the adsorber module, an adsorber apparatus for whole blood treatment as shown in Fig. 4 is constructed.

**[0074]** A blood from a patient suffering from rheumatoid arthritis is heparinized. An aliquot of the blood is subjected to measurement of amounts of rheumatoid factor, immune complex and platelet. The measurement of the amount of rheumatoid factor is conducted by a passive hemagglutination test method using a RAHA test kit (manufactured and sold by Fuji Zoki Pharmaceutical Co., Ltd., Japan), and the measurement of the amount of immune complex is conducted by a Raji Cell method as described in "Immune Complex Disease", 1st ed., published by Ishiyaku Publishers, Inc., Tokyo, Japan (1982), pp. 93-94. The measurement of the amount of platelet is conducted by a well known Brecher-Cronkite method in which a blood sample is diluted 100 times with a 1 % aqueous ammonium oxalate solution and subjected to measurement of the platelet concentration by using a hemocytometer. The blood is introduced to the adsorber apparatus from the patient (donor) through blood inlet 9 and flowed to adsorber module 13 at a flow rate of 50 ml/min through conduits 10 and 12 by means of pump 11 (blood transport means). In adsorber module 13, the blood is allowed to flow through the hollows of hollow fibers 1, and the plasma of the blood is allowed to pass through the membrane of each hollow fiber. The thus separated plasma is flowed to conduit 20 through a bypass, i.e., through conduits 16 and 18 by means of pump 17 at a flow rate of one third of the flow rate of the blood flowed into adsorber module 13 by means of pump 11. On the other hand, the cell-enriched blood which has not been passed through the bypass is flowed to conduit 20 through conduit 19. In conduit 20, the plasma is combined with the cell-enriched blood and the resultant mixture is returned to the donor through blood outlet 15. The blood from the donor is recycled through the same adsorber apparatus. The recycling is continued for 30 min. With this adsorber apparatus, recycling of the blood is stably, sufficiently

performed using an extracorporeal circuit blood volume as small as 210 ml. This volume is satisfactorily small from the viewpoint of attaining the whole blood treatment with a decreased extracorporeal circuit blood volume being taken outside the body.

**[0075]** The blood treatment by the adsorber apparatus can be stably conducted without blood coagulation and hemolysis.

**[0076]** After the blood recycling for 30 min, an aliquot of the recycled blood is collected from blood outlet 20 and the amounts of rheumatoid factor, immune complex and platelet in the blood collected are measured in the same manner as mentioned above.

**[0077]** As a result, it is found that the amounts of the rheumatoid arthritis and immune complex before the blood treatment which are 1280 and 120  $\mu\text{g/ml}$ , respectively, are decreased to 320 and 28  $\mu\text{g/ml}$ , respectively, after the blood treatment.

**[0078]** On the other hand, the platelet concentration before the treatment is 340,000 cells/ $\text{mm}^3$ , while the platelet concentration after the treatment is 300,000 cells/ $\text{mm}^3$ . That is, the platelet concentration is not so lowered by the whole blood treatment.

**[0079]** As apparent from the above, by the use of the adsorber apparatus of the present invention, malignant substances (i.e., rheumatoid factor and immune complex) are selectively removed with little loss of platelets.

#### Example 2

**[0080]** Epoxy-activated hollow fibers are prepared in the same manner as in Example 1, and immersed in a 30 % (w/w) aqueous dextran sulfate solution (pH 13) and kept at 50 °C for 24 hours, to thereby link dextran sulfate (a ligand) to the overall surface of each hollow fiber. The resultant hollow fibers are sufficiently washed with water. Using the washed hollow fibers, an adsorber module is prepared in the same manner as in Example 1.

**[0081]** Then, using the thus obtained adsorber module, an adsorber apparatus for blood treatment is constructed as shown in Fig. 3.

**[0082]** A blood from a patient suffering from familial hypercholesterolemia and having a hematocrit (Ht) of 35 % is heparinized. An aliquot of the blood is subjected to measurement of the total cholesterol concentration and platelet concentration. The measurement of total cholesterol concentration is conducted by an enzyme assay using a test kit Cholesterol C-Test Wako, manufactured and sold by Wako Pure Chemicals Industries Ltd., Japan. The most part of cholesterol present in the blood of a patient of familial hypercholesterolemia is derived from low-density lipoproteins, and the term "total cholesterol" means to include cholesterol and low-density lipoproteins. The measurement of platelet concentration is conducted in the same manner as in Example 1. The blood is introduced to the adsorber apparatus from the patient (donor) through blood inlet 9 and flowed to adsorber module 13 at a flow rate of 50 ml/min through conduits 10 and 12 by means of pump 11. 10 Minutes later, it is observed that the interstices extending between the casing and the hollow fibers of the adsorber module and between the outer surfaces of individual hollow fibers have been filled with plasma assuming light yellow. The blood treated with the adsorber module is returned to the donor through blood outlet 15. The blood from the donor is recycled through the same adsorber apparatus in the same manner as in Example 1. The recycling is continued for 30 minutes. With this adsorber apparatus, recycling of the blood is stably, sufficiently performed using an extracorporeal circuit blood volume as small as 160 ml. This volume is satisfactorily small from the viewpoint of attaining the whole blood treatment with a decreased extracorporeal circuit blood volume being taken outside the body. 30 Minutes later, an aliquot of the recycled blood is collected from blood outlet 15 and the total cholesterol concentration of the collected blood is measured according to the same method as mentioned above. As a result, it is found that the total cholesterol concentration is decreased to 110 mg/dl after the blood treatment, which is extremely low as compared to the total cholesterol concentration of the blood before the blood treatment, that is, 540 mg/dl.

**[0083]** On the other hand, the platelet concentration of the blood is also determined after the blood treatment. As a result, it is found that the platelet concentration after the blood treatment is 250,000 cells/ $\text{mm}^3$ , while the platelet concentration before the blood treatment is 270,000 cells/ $\text{mm}^3$ . That is, the platelet concentration is not so lowered by the blood treatment.

**[0084]** As apparent from the above, by the use of the adsorber apparatus of the present invention, a malignant substances (i.e., cholesterol and low-density lipoproteins) are selectively removed with little loss of platelets.

#### Examples 3 and 4 and comparative example 1

**[0085]** A high-density polyethylene (HI-ZEX 2208J, a product of Mitsui Petrochemical Co., Japan) having a density of 0.968 g/ $\text{cm}^3$  and a melt index, as measured in accordance with ASTM D1238, of 5.5 is extruded through a spinning nozzle having an annular orifice outside diameter of 33 mm and an annular orifice inside diameter of 25 mm at a winding temperature of 150 °C, and at a polymer extrusion rate of 16 g/min and a winding rate as shown in Table 1. The thus

obtained hollow fiber is subjected to annealing at 115 °C for 2 hours. The annealed hollow fiber is then cold-stretched at room temperature at a stretching ratio (times) (the ratio of the length of the stretched hollow fiber to the length of the hollow fiber before stretching, expressed by times) of 1.33 times by passing the hollow fiber through stretching rolls arranged to provide a stretching-path of 200 mm. Then, the cold-stretched hollow fiber is hot-stretched successively at 78 °C, 95 °C and 98 °C to thereby stretch the hollow fiber at a ratio (%) of the length of the hollow fiber to the original length of the hollow fiber before cold-stretching and hot-stretching, of 480 %. The resultant hollow fiber is heat set at 115 °C for 2 min, to thereby obtain a porous polyethylene hollow fiber.

**[0086]** The polyethylene hollow fiber is subjected successively to polyvinyl alcohol-treatment, epoxy-activation, and treatment for linking ligands in the same manner as in Example 1. Then, an adsorber module for blood treatment is prepared using the porous polyethylene hollow fiber in the same manner as in Example 1. The average inside diameter (D), the average effective length (L) and the total surface area of the hollow fibers in the adsorber module are shown in Table 1. Using the adsorber module, substantially the same adsorber apparatus for blood treatment as shown in Fig. 4 except that a plasma circuit including pump 17 and conduit 18 is removed, and a container is attached to conduit 16 in order to collect plasma separated by adsorber module 13 is constructed.

**[0087]** A blood from a patient (donor) suffering from rheumatoid arthritis is heparinized and introduced to the adsorber apparatus through blood inlet 9. The introduced blood is flowed to adsorber module 13 through conduits 10 and 12 by means of pump 11, and plasma separated by adsorber module 13 is collected through plasma outlet 16. With this adsorber apparatus, recycling of the blood is stably, sufficiently performed using an extracorporeal circuit blood volume as small as 210 ml. This volume is satisfactorily small from the viewpoint of attaining the whole blood treatment with a decreased extracorporeal circuit blood volume being taken outside the body. The amount of the collected plasma is measured. Further, the rheumatoid factor concentration of the collected plasma is determined in the same manner as in Example 1. The results are shown in Table 1.

**[0088]** As apparent from the results shown in Table 1, although the average inside diameters of the individual hollow fibers are different between the hollow fibers used in Examples 3 to 5, the surface areas of the hollow fibers in the prepared adsorber modules of Examples 3 to 5 are almost the same. However, the larger the  $L/D^2$  value, the smaller the amount of rheumatoid factor in the treated blood becomes. On the other hand, the larger the  $L/D^2$  value, the larger the collection rate of plasma becomes. This means that the smaller the inside diameter of the hollow fiber, the higher the efficiency of plasma separation becomes. Particularly, when the  $L/D^2$  value is  $2,000 \text{ mm}^{-1}$  or higher as in Examples 3 and 4, plasma separation can be conducted with extremely high efficiency.

Table 1

Example	Winding rate (m/min)	Average inside diameter of hollow fiber (D, mm)	Average effective length of hollow fiber (L, mm)	$L/D^2 \text{ (mm}^{-1}\text{)}$	Surface area of hollow fibers in adsorber module ( $\text{m}^2$ )	Separation rate of plasma (ml/min)	Amount of rheumatoid factor after the blood treatment
3	300	0.340	250	2,200	110	21	160
4	600	0.280	250	3,200	100	27	80
comp. ex. 1	200	0.380	250	1,700	120	11	640
Condition: Blood flow rate is 50 ml/min (Ht=35%) and the original amount of rheumatoid factor before the blood treatment is 1280.							

## Comparative Example 2

**[0089]** An adsorber material comprising a granular carrier and tryptophan linked to the carrier is used for the adsorption of rheumatoid factor. The adsorber material is produced as follows.

**[0090]** A uniform mixture of 100 g of vinyl acetate, 64.3 g of triallyl isocyanurate, 100 g of ethyl acetate, 100 g of heptane, 7.5 g of polyvinyl acetate (degree of polymerization: 500) and 3.8 g of 2,2'-azobisisobutyronitrile, and 400 ml of an aqueous solution containing 1 % by weight of polyvinyl alcohol, 0.05 % by weight of sodium dihydrogenphosphate dihydrate and 1.5 % by weight of sodium dihydrogenphosphate dodecahydrate are charged in a flask and sufficiently mixed by stirring. Then, the resultant mixture is heated, while stirring, successively at 65 °C for 18 hours and at 75 °C for 5 hours to conduct a suspension polymerization. The thus obtained granular copolymer is filtered off and washed

with water. The thus obtained copolymer is subjected to acetone extraction. The resultant copolymer is subjected to ester exchange reaction at 40 °C for 18 hours in a solution of 46.5 g of sodium hydroxide in 2 ℓ of methanol. The thus obtained gel of the copolymer has an average particle diameter of 100 μm, a vinyl alcohol unit (qOH) per unit weight of 8.9 meq/g, a specific surface area of 65 m<sup>2</sup>/g and a cut-off molecular weight of 9 x10<sup>5</sup> as measured using a dextran.

5 [0091] Then, 50 g of the obtained gel (dry basis) is suspended in 600 ml of dimethyl sulfoxide and to the resultant suspension are added 391.5 ml of epichlorohydrin and 50 ml of 30 % (w/w) aqueous sodium hydroxide solution. The mixture is stirred at 30 °C for 5 hours to activate the gel.

[0092] After the gel activation, the resultant gel is washed successively with dimethylsulfoxide and with water, and suction-dehydrated. The thus obtained activated gel is suspended in 800 ml of 0.1 M sodium carbonate buffer contain-  
10 ing 8.15 g of tryptophan. The suspension is stirred at 50 °C for 14 hours to link the tryptophan to the overall surface of the gel. Thereafter, to the mixture is added 165 ml of an aqueous tris(hydroxyethyl)aminomethane solution having a tris(hydroxyethyl)aminomethane concentration of 60.6 mg/ml. Then, the resultant mixture is heated at 50 °C for 5 hours while stirring so that the active groups of the gel remaining unlinked by tryptophan are blocked. The resultant gel is suf-  
15 ficiently washed with water, to thereby obtain an adsorber material. The amount of the tryptophan linked to the adsorber material is 360 μmol/g (dry basis).

[0093] The adsorber material is filled in a column having an inside diameter of 40 mm and a length of 318 mm to obtain an adsorber column. Using the adsorber column, an adsorber apparatus as shown in Fig. 5 is constructed. In Fig. 5, numeral 24 designates the adsorber column and numeral 21 designates a plasma separator. The adsorber col-  
20 umn 24 is connected to the plasma outlet of plasma separator 21 through conduit 21, pump 17 and conduit 23 so that the plasma separated by plasma separator 21 can be conveyed to adsorber column 24 by means of pump 17. As the plasma separator, Plasmaflo<sup>®</sup> AP05H manufactured and sold by Asahi Medical Co., Ltd., Japan is used.

[0094] A blood from a patient suffering from rheumatoid arthritis is heparinized. An aliquot of the blood is subjected to measurement of amount of rheumatoid factor by a passive hemagglutination test method using a RAHA test kit (man-  
25 ufactured and sold by Fuji Zoki Pharmaceutical Co., Ltd., Japan). The blood is introduced to the adsorber apparatus from the patient (donor) through blood inlet 9 and flowed to plasma separator 21 at a flow rate of 50 ml/min through con-  
30 duits 10 and 12 by means of pump 11. The blood is separated into plasma and cell-enriched blood by plasma separator 21, and the plasma is flowed to adsorber column 24 through conduits 22 and 23 by means of pump 17 at a flow rate of 17 ml/min. In conduit 27, the plasma treated with adsorber column 24 and flowed through conduit 25 is combined with cell-enriched blood flowed through conduit 26, and the resultant mixture is returned to the donor through blood outlet  
30 15. The blood from the donor is recycled through the same adsorber apparatus. The recycling is continued for 30 min.

[0095] After the blood recycling for 30 min, an aliquot of the recycled blood is collected from blood outlet 27 and the amount of rheumatoid factor in the blood is measured in the same manner as in Example 1.

[0096] As a result, it is found that the amount of rheumatoid factor before the blood treatment which is 1280, is decreased to 160 after the blood treatment, which is comparable to the amount of the rheumatoid factor in the blood  
35 after the blood treatment conducted in Example 3. However, with this adsorber apparatus, recycling of the blood is per-  
formed using an extracorporeal circuit blood volume as large as 610 ml. This volume is disadvantageously very large so that a heavy burden is posed on the patient during the blood treatment.

## Claims

40 1. An adsorber module for removing malignant components of whole blood by adsorption, comprising:

a casing provided with a blood introduction means (2) and a blood withdrawal means (3), and  
45 a plurality of porous hollow fibers (1) substantially equal in length arranged substantially in parallel relationship and bonded together at both end portions thereof to form a bundle, each porous hollow fiber of said bundle having openings at both terminal ends thereof,  
said bundle being disposed in said casing along the length of said casing,  
said both end portions of the hollow fibers of said bundle being fluid-tightly connected to said blood introduction means and said blood withdrawal means, respectively, thereby establishing communication between said  
50 blood introduction means and said blood withdrawal means through said bundle of hollow fibers,  
each porous hollow fiber comprising a membranous porous resin matrix having pores therewithin and open-  
ings on both surfaces thereof, said pores cooperating with said openings to form throughpaths running between both the surfaces of said resin matrix, and a plurality of ligands (8) attached to the overall surface of  
said porous resin matrix,  
55 each of said ligands being capable of selectively interacting with the malignant components to be adsorbed on said hollow fibers,  
said ligands having low antigenicity such that when the ligands are in contact with blood corpuscles, the lig-  
ands do not cause hemolysis of erythrocyte, sensitization of leukocyte, or adherence and/or aggregation of

platelet,

said porous resin matrix having an average pore diameter of from 0.005 to 3  $\mu\text{m}$ ,

said porous hollow fibers having an average effective length (L mm) and an average inner diameter (D mm) which satisfy the inequalities

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000, \text{ and } 150 \mu\text{m} \leq D \leq 400 \mu\text{m}$$

said average effective length being defined as an average of the lengths of said porous hollow fibers minus the lengths of said both end portions of said porous hollow fibers at which the fibers are bonded together and fluid-tightly connected to said whole blood introduction means and said blood withdrawal means, respectively.

2. The module according to claim 1, wherein said ligand is linked to the overall surface of said porous resin matrix by covalent bonding.

3. The module according to any one of claims 1 or 2, wherein said ligand is a hydrophobic compound.

4. The module according to any one of claims 1 to 3, wherein said ligand is a polyanion.

5. The module according to any one of claims 1 to 4, wherein said average pore diameter is in the range of from 0.01 to 2  $\mu\text{m}$ .

6. The module according to any one of claims 1 to 5, wherein said casing is further provided with at least one opening (4) for withdrawing the plasma.

7. The module according to any one of claims 1 to 6, wherein said resin matrix is composed of at least one resin selected from the group consisting of cellulose, a cellulose derivative, a water-insoluble polyvinyl alcohol, a copolymer of ethylene and vinyl alcohol, a polyolefin, a polysulfone and a polytetrafluoroethylene.

8. An adsorber apparatus for removing malignant components of whole blood by adsorption, comprising:

(a) an adsorber module (13,) comprising:

a casing provided with a blood introduction means (2) and a blood withdrawal means (3), and a plurality of porous hollow fibers (1) substantially equal in length arranged substantially in parallel relationship and bonded together at both end portions thereof to form a bundle, each porous hollow fiber of said bundle having openings at both terminal ends thereof,

said bundle being disposed in said casing along the length of said casing, said both end portions of the hollow fibers of said bundle being fluid-tightly connected to said blood introduction means and said blood withdrawal means, respectively, thereby establishing communication between said blood introduction means and said blood withdrawal means through said bundle of hollow fibers,

each porous hollow fiber comprising a membranous porous resin matrix having pores therewithin and openings on both surfaces thereof, said pores cooperating with said openings to form throughpaths running between both the surfaces of said resin matrix, and a plurality of ligands attached to the overall surface of said porous resin matrix,

each of said ligands being capable of selectively interacting with the malignant components to be adsorbed on said hollow fibers,

said ligands having low antigenicity such that when the ligands are in contact with blood corpuscles, the ligands do not cause hemolysis of erythrocyte, sensitization of leukocyte, or adherence and/or aggregation of platelet,

said porous resin matrix having an average pore diameter of from 0.005 to 3  $\mu\text{m}$ ,

said porous hollow fibers having an average effective length (L mm) and an average inner diameter (D mm) which satisfy the inequalities:

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000, \text{ and } 150 \mu\text{m} \leq D \leq 400 \mu\text{m}$$

said average effective length being defined as an average of the lengths of said porous hollow fibers minus the lengths of said both end portions of said porous hollow fibers at which the fibers are bonded together

and fluid-tightly connected to said whole blood introduction means and said blood withdrawal means, respectively,

(b) a blood introducing passage means comprising a first conduit having one end fluid-tightly connected to said blood introduction means of module (a), a second conduit having a blood inlet at one end thereof, and a blood transport means disposed between and fluid-tightly connected to the other ends of said first and second conduits; and

(c) a blood withdrawing passage means comprising a third conduit having one end fluid-tightly connected to said blood withdrawal means of module (a) and having a blood outlet at the other end thereof.

9. The apparatus according to claim 8, comprising an adsorber module according to any of the claims 2 to 7.

10. The apparatus according to claim 9, wherein said casing of module (a) is further provided with at least one opening (4) for withdrawing the plasma,

said opening being fluid-tightly connected to said blood withdrawing passage means through a bypass comprising a fourth conduit (16,22) having one end fluid-tightly connected to said opening, a fifth conduit (18; 23,25) having one end fluid-tightly connected to said blood withdrawing passage at its intermediate position, and a plasma transport means (17) disposed between and fluid-tightly connected to the other ends of said fourth and fifth conduits.

## Patentansprüche

1. Adsorbermodul zum Entfernen von bösartigen Vollblutbestandteilen durch Adsorption, das umfaßt:

ein Gehäuse, das mit einer Blut-Zuleitungseinrichtung (2) und einer Blut-Entnahmeeinrichtung (3) versehen ist, und

eine Vielzahl von porösen Hohlfasern (1) von im wesentlichen gleicher Länge, die im wesentlichen parallel zueinander angeordnet und an ihren beiden Endteilen miteinander unter Bildung eines Bündels verbunden sind, wobei jede poröse Hohlaser des Bündels Öffnungen an ihren beiden Enden hat,

wobei dieses Bündel in dem Gehäuse in Richtung der Länge des Gehäuses angeordnet ist,

diese beiden Endteile der Hohlfasern des Bündels flüssigkeitsdicht mit der Blut-Zuleitungseinrichtung und der Blut-Entnahmeeinrichtung verbunden sind, so daß auf diese Weise eine Verbindung zwischen der Blut-Zuleitungseinrichtung und der Blut-Entnahmeeinrichtung durch das Bündel aus Hohlfasern ausgebildet wird,

wobei jede poröse Hohlaser ein membranförmige poröse Harzmatrix umfaßt, die darin befindliche Poren und Öffnungen an ihren beiden Oberflächen hat, wobei die Poren mit den Öffnungen unter Bildung von Durchgängen zusammenwirken, welche zwischen den beiden Oberflächen der Harzmatrix verlaufen, und mehrere Liganden (8) aufweist, die an der gesamten Oberfläche der porösen Harzmatrix gebunden sind,

wobei diese Liganden zur selektiven Wechselwirkung mit den auf den Hohlfasern zu adsorbierenden bösartigen Bestandteilen befähigt sind,

diese Liganden niedere Antigenizität haben, so daß, wenn die Liganden in Kontakt mit Blutkörperchen stehen, diese Liganden nicht die Hämolyse von Erythrocyten, Sensibilisierung von Leucocyten oder ein Aneinanderhaften und/oder eine Aggregation von Blutplättchen bewirken,

die poröse Harzmatrix einen durchschnittlichen Porendurchmesser von 0,005 bis 3 µm hat und

die porösen Hohlfasern eine durchschnittliche wirksame Länge (L mm) und einen durchschnittlichen Innendurchmesser (D mm) haben, welche die Ungleichungen

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000 \text{ und } 150 \text{ µm} \leq D \leq 400 \text{ µm}$$

erfüllen, wobei die durchschnittliche wirksame Länge definiert ist als der Durchschnitt der Längen der porösen Hohlfasern minus die Längen der beiden Endteile der porösen Hohlfasern, an denen die Fasern miteinander verbunden sind und flüssigkeitsdicht mit der Zuleitungseinrichtung für das Vollblut und mit der Blut-Entnahmeeinrichtung verbunden sind.

2. Modul gemäß Anspruch 1, worin der Ligand durch kovalente Bindung an die gesamte Oberfläche der porösen Harzmatrix gebunden ist.

3. Modul gemäß einem der Ansprüche 1 oder 2, bei dem der Ligand eine hydrophobe Verbindung ist.

4. Modul gemäß einem der Ansprüche 1 bis 3, bei dem der Ligand ein Polyanion ist.
5. Modul gemäß einem der Ansprüche 1 bis 4, bei dem der durchschnittliche Porendurchmesser im Bereich von 0,01 bis 2 µm ist.
6. Modul gemäß einem der Ansprüche 1 bis 5, bei dem das Gehäuse außerdem mindestens eine Öffnung (4) zur Entnahme von Plasma aufweist.
7. Modul gemäß einem der Ansprüche 1 bis 6, bei dem die Harzmatrix aus mindestens einem Harz besteht, das aus der aus Cellulose, einem Cellulosederivat, einem wasserunlöslichen Polyvinylalkohol, einem Ethylen-Vinylalkohol-Copolymeren, einem Polyolefin, einem Polysulfon und einem Polytetrafluorethylen bestehenden Gruppe ausgewählt ist.
8. Adsorbervorrichtung zur Entfernung von bösartigen Vollblutbestandteilen durch Adsorption, das umfaßt:  

ein Gehäuse, das mit einer Blut-Zuleitungseinrichtung (2) und einer Blut-Entnahmeeinrichtung (3) versehen ist, und  
eine Vielzahl von porösen Hohlfasern (1) von im wesentlichen gleicher Länge, die im wesentlichen parallel zueinander angeordnet und an ihren beiden Endteilen miteinander unter Bildung eines Bündels verbunden sind, wobei jede poröse Hohlaser des Bündels Öffnungen an ihren beiden Enden hat, wobei dieses Bündel in dem Gehäuse in Richtung der Länge des Gehäuses angeordnet ist, diese beiden Endteile der Hohlfasern des Bündels flüssigkeitsdicht mit der Blut-Zuleitungseinrichtung und der Blut-Entnahmeeinrichtung verbunden sind, so daß auf diese Weise eine Verbindung zwischen der Blut-Zuleitungseinrichtung und der Blut-Entnahmeeinrichtung durch das Bündel aus Hohlfasern ausgebildet wird, wobei jede poröse Hohlaser ein membranförmige poröse Harzmatrix umfaßt, die darin befindliche Poren und Öffnungen an ihren beiden Oberflächen hat, wobei die Poren mit den Öffnungen unter Bildung von Durchgängen zusammenwirken, welche zwischen den beiden Oberflächen der Harzmatrix verlaufen, und mehrere Liganden (8) aufweist, die an der gesamten Oberfläche der porösen Harzmatrix gebunden sind, wobei diese Liganden zur selektiven Wechselwirkung mit den auf den Hohlfasern zu adsorbierenden bösartigen Bestandteilen befähigt sind, diese Liganden niedere Antigenizität haben, so daß, wenn die Liganden in Kontakt mit Blutkörperchen stehen, diese Liganden nicht die Hämolyse von Erythrocyten, Sensibilisierung von Leucocyten oder ein Aneinanderhaften und/oder eine Aggregation von Blutplättchen bewirken, die poröse Harzmatrix einen durchschnittlichen Porendurchmesser von 0,005 bis 3 µm hat und die porösen Hohlfasern eine durchschnittliche wirksame Länge (L mm) und einen durchschnittlichen Innendurchmesser (D mm) haben, welche die Ungleichungen

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000 \text{ und } 150 \text{ µm} \leq D \leq 400 \text{ µm}$$

erfüllen, wobei die durchschnittliche wirksame Länge definiert ist als der Durchschnitt der Längen der porösen Hohlfasern minus die Längen der beiden Endteile der porösen Hohlfasern, an denen die Fasern miteinander verbunden sind und flüssigkeitsdicht mit der Zuleitungseinrichtung für das Vollblut und mit der Blut-Entnahmeeinrichtung verbunden sind,

(b) eine Blutzuführungs-Durchleiteeinrichtung, die eine erste Leitung (12), deren eines Ende flüssigkeitsdicht mit der Blut-Zuleitungseinrichtung des Moduls (a) verbunden ist, eine zweite Leitung (10), an deren einen Ende ein Bluteinlaß (9) vorgesehen ist und eine Bluttransporteinrichtung (11), die zwischen den anderen Enden der ersten und der zweiten Leitung angeordnet und mit diesen flüssigkeitsdicht verbunden ist, aufweist und

(c) eine Blutentnahme-Durchleiteeinrichtung, die eine dritte Leitung (14, 19, 20, 26, 27) aufweist, deren eines Ende flüssigkeitsdicht mit der Blut-Entnahmeeinrichtung des Moduls (a) verbunden ist und die eine Blut-Auslaßöffnung (15) an ihrem anderen Ende hat.
9. Vorrichtung gemäß Anspruch 8, enthaltend ein Adsorbermodul gemäß einem der Ansprüche 1 bis 7.
10. Vorrichtung gemäß Anspruch 9, in der das Gehäuse von Modul (a) außerdem mit mindestens einer Öffnung (4) zur Entnahme des Plasmas versehen ist,



wobei diese Öffnung flüssigkeitsdicht über eine Umgehungsleitung mit einer vierten Leitung (16, 22), deren eines Ende flüssigkeitsdicht mit dieser Öffnung verbunden ist, mit der Blutentnahme-Durchleiteeinrichtung verbunden ist, einer fünften Leitung (18, 23, 25), deren eines Ende flüssigkeitsdicht mit der Blutentnahme-Durchleiteeinrichtung an einer Stelle in deren Mitte verbunden ist und mit einer Plasma-Transporteinrichtung (17) versehen ist, die zwischen den anderen Enden der vierten und der fünften Leitung angeordnet und mit diesen flüssigkeitsdicht verbunden ist.

## Revendications

1. Module adsorbeur pour éliminer les constituants malins du sang total, qui comprend :

un boîtier muni d'un moyen (2) d'introduction du sang et d'un moyen (3) de soutirage du sang, et une pluralité de fibres (1) tubulaires poreuses de longueur sensiblement égale, disposées d'une manière sensiblement parallèle et liées les unes aux autres en leurs deux parties d'extrémité pour former un faisceau, chaque fibre tubulaire poreuse de ce faisceau comportant des ouvertures à ses deux extrémités terminales, le faisceau étant disposé dans le boîtier suivant la longueur du boîtier, les deux parties d'extrémité des fibres tubulaires du faisceau communiquant, d'une manière étanche aux fluides, respectivement avec le moyen d'introduction du sang et le moyen de soutirage du sang, de façon à établir une communication entre le moyen d'introduction du sang et le moyen de soutirage du sang, en passant par le faisceau de fibres tubulaires, chaque fibre tubulaire poreuse comprenant une matrice de résine poreuse membranaire ayant des pores en son sein et des ouvertures sur ses deux faces, les pores coopérant avec les ouvertures pour former des trajets allant entre les deux faces de la matrice de résine et une pluralité de ligands (8) fixés à toute la surface de la matrice de résine poreuse, chacun des ligands pouvant interagir avec les constituants malins à adsorber sur les fibres tubulaires, les ligands ayant une faible antigénicité, telle que, lorsque les ligands sont en contact avec des corpuscules de sang, les ligands ne provoquent pas d'hémolyse d'érythrocyte, de sensibilisation de leucocyte ni d'adhérence et/ou d'aggrégation de plaquette, la matrice de résine poreuse ayant un diamètre moyen de pores de 0,005 à 3 µm, les fibres tubulaires poreuses ayant une longueur effective moyenne (L mm) et un diamètre intérieur moyen (D mm) qui satisfont à l'inégalité suivante :

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000 \text{ et } 150 \text{ µm} \leq D \leq 400 \text{ µm}$$

la longueur effective moyenne étant définie par la moyenne des fibres tubulaires poreuses, diminuée de la longueur des deux parties d'extrémité des fibres tubulaires poreuses où les fibres sont liées les unes aux autres et communiquent d'une manière étanche aux fluides respectivement avec le moyen d'introduction du sang et avec le moyen de soutirage du sang.

2. Module suivant la revendication 1, dans lequel le ligand est lié à toute la surface de la matrice en résine poreuse par liaison covalente.
3. Module suivant l'une quelconque des revendications 1 ou 2, dans lequel le ligand est un composé hydrophobe.
4. Module suivant l'une quelconque des revendications 1 à 3, dans lequel le ligand est un polyanion.
5. Module suivant l'une quelconque des revendications 1 à 4, dans lequel le diamètre moyen de pores est compris entre 0,01 et 2 µm.
6. Module suivant l'une quelconque des revendications 1 à 5, dans lequel le boîtier est muni en outre d'au moins une ouverture (4) pour enlever le plasma.
7. Module suivant l'une quelconque des revendications 1 à 6, dans lequel la matrice en résine est composée d'au moins une résine choisie parmi la cellulose, un dérivé de cellulose, de l'alcool polyvinylique insoluble dans l'eau, un copolymère d'éthylène et d'alcool vinylique, une polyoléfine, une polysulfone et un polytétrafluoroéthylène.
8. Appareillage d'adsorption pour éliminer des constituants malins du sang entier comprenant :

(a) un module (13) d'adsorbeur comprenant :

un boîtier muni de moyens (2) d'introduction du sang et de moyens (3) de soutirage du sang, et une pluralité de fibres (1) tubulaires poreuses de longueur sensiblement égale disposées sensiblement parallèlement et liées ensemble à leurs deux parties d'extrémité pour former un faisceau, chaque fibre tubulaire poreuse du faisceau ayant des ouvertures à ses deux extrémités, le faisceau étant disposé dans le boîtier suivant la longueur du boîtier, les deux parties d'extrémité des fibres tubulaires du faisceau communiquant d'une manière étanche aux fluides avec les moyens d'introduction du sang et avec des moyens de soutirage du sang respectivement de manière à établir une communication entre les moyens d'introduction du sang et les moyens de soutirage du sang par le faisceau de fibres tubulaires, chaque fibre tubulaire poreuse comprenant une matrice en résine poreuse membranaire ayant des pores en son sein et des ouvertures sur ses deux faces, les pores coopérant avec les ouvertures pour former des trajets allant entre les deux faces de la matrice de résine et une pluralité de ligands (8) fixés à toute la surface de la matrice de résine poreuse, chacun des ligands étant susceptible d'interagir sur les constituants malins à adsorber sur les fibres tubulaires, le ligands ayant une faible antigénicité, telle que, lorsque les ligands sont en contact avec des corpuscules de sang, ils ne provoquent pas d'hémolyse d'érythrocyte, de sensibilisation de leucocyte, ni d'adhérence et/ou d'aggrégation de plaquette. la matrice en résine poreuse ayant un diamètre moyen de force de 0,005 à 3  $\mu\text{m}$ , les fibres tubulaires poreuses ayant une longueur effective moyenne (L mm) et un diamètre intérieur moyen (D mm) qui satisfont aux inégalités

$$L/D^2 \text{ (mm}^{-1}\text{)} \geq 2000 \text{ et } 150 \mu\text{m} \leq D \leq 400 \mu\text{m}$$

la longueur effective moyenne étant définie comme la moyenne des longueurs des fibres tubulaires poreuses, déduction faite des longueurs des deux parties d'extrémité des fibres tubulaires poreuses où les fibres sont liées ensemble et communiquent d'une manière étanche aux gaz avec les moyens d'introduction du sang et avec les moyens de soutirage du sang respectivement,

(b) un passage d'introduction du sang comprenant un premier conduit dont une extrémité communique de manière étanche au fluide avec les moyens d'introduction du sang du module (a), un deuxième conduit ayant une entrée (9) pour le sang à l'une de ses extrémités et des moyens de transport du sang interposés entre les autres extrémités des premier et deuxième conduits et communiquant avec eux de manière étanche aux fluides; et

(c) un passage de soutirage du sang comprenant un troisième conduit dont une extrémité communique d'une manière étanche aux fluides avec les moyens de soutirage du sang du module (a) et ayant une sortie pour le sang à l'autre extrémité.

**9.** Appareillage suivant la revendication 8, comprenant un module d'adsorbeur suivant l'une quelconque des revendications 2 à 7.

**10.** Installation suivant la revendication 9, dans laquelle le boîtier du module (a) est muni en outre d'au moins une ouverture (4) de soutirage du plasma,

l'ouverture communiquant d'une manière étanche aux fluides avec le passage de soutirage du sang par une dérivation comprenant un quatrième conduit (16,22) dont une extrémité communique d'une manière étanche aux fluides avec l'ouverture, un cinquième conduit (18,23,25) dont une extrémité communique d'une manière étanche aux fluides avec la passage de soutirage du sang en sa position intermédiaire et des moyens (17) de transport du plasma interposés entre les autres extrémités des quatrième et cinquième conduits et communiquant avec ceux-ci d'une manière étanche aux fluides.

FIG. 1

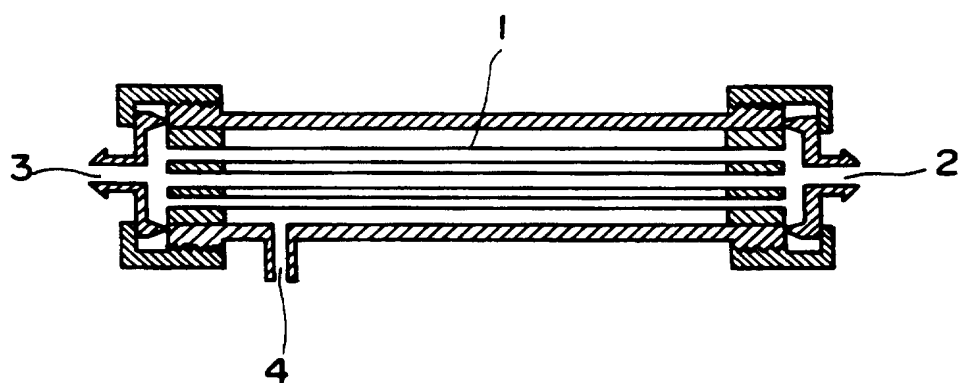


FIG. 2

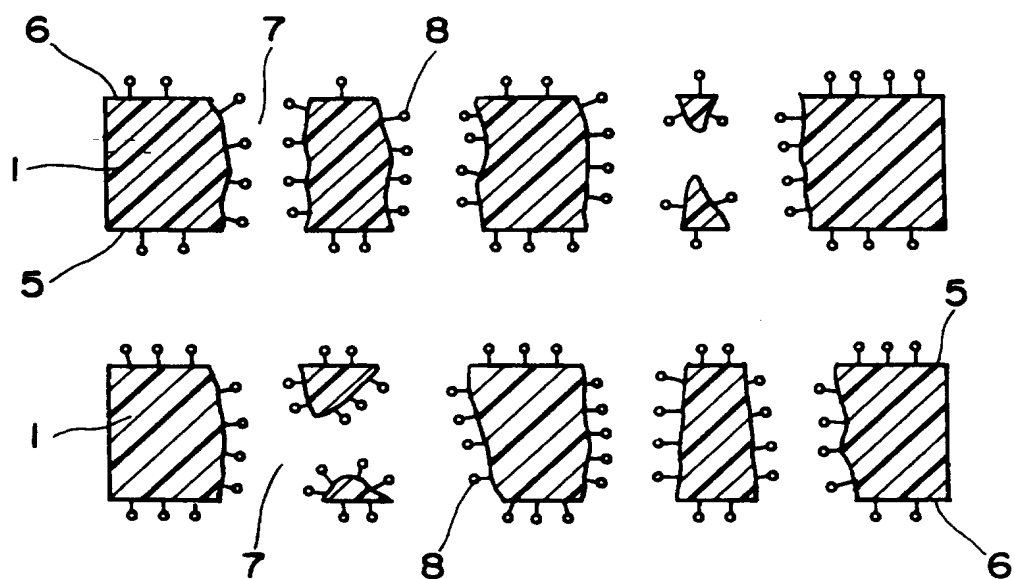


FIG. 3

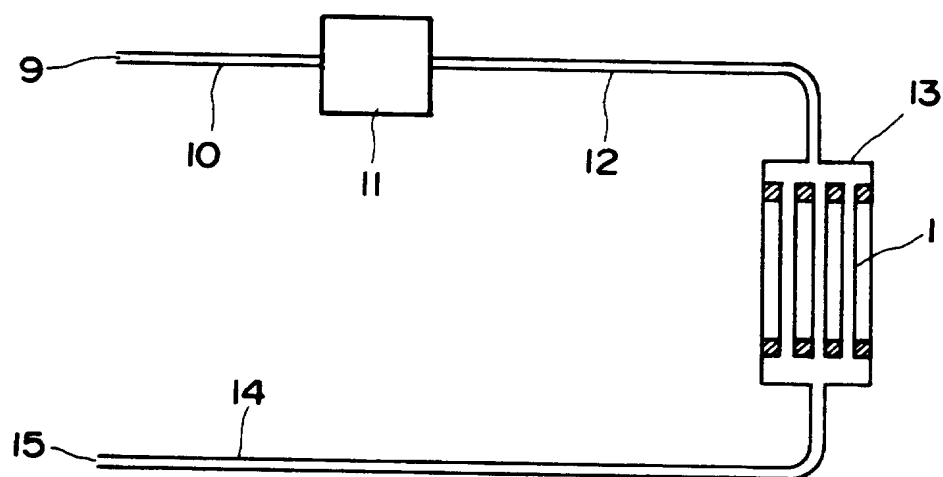


FIG. 4

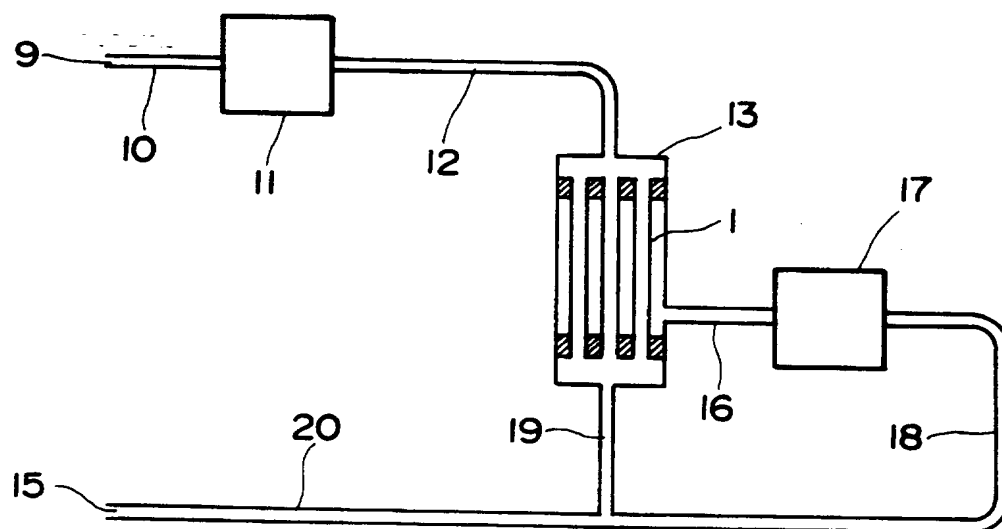


FIG. 5

